## A Stereocontrolled Synthesis of a C<sub>19</sub>-C<sub>32</sub> / C<sub>17</sub>-C<sub>30</sub> Segment for Swinholide A and Misakinolide A, Cytotoxic Dimeric Macrolides from *Theonella Swinhoei*.

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Abstract: The C<sub>19</sub>-C<sub>32</sub> / C<sub>17</sub>-C<sub>30</sub> segment (-)-5 of swinholide A / misakinolide A was prepared in 15 steps (6% yield) from  $(\pm)$ -13. Key steps include the Sharpless epoxidation,  $13 \rightarrow 14$ , the acetal allylation,  $12 \rightarrow 16$ , the *anti* aldol,  $17 + 11 \rightarrow 9$ , and the alkene hydroboration,  $19 \rightarrow 20$ .

Swinholide A, a novel cytotoxic macrolide isolated from marine sponges of the genus *Theonella* swinhoei, was first reported by Carmely and Kashman in 1985.<sup>1</sup> While originally misassigned as a monomeric macrolide,<sup>1</sup> more recent mass spectroscopic<sup>2a</sup> and X-ray crystallographic<sup>2b-d</sup> studies showed it to be a symmetrical dimer having the 44-membered dilactone structure 1 (Scheme 1). Several other dimeric macrolides have also been isolated from *Theonella*, including the desmethyl analogues swinholides B (2) and C (3),<sup>2e</sup> and the closely related 40-membered dilactone, misakinolide A (4)<sup>3a-c</sup> (= bistheonellide A<sup>3b,d</sup>). These are all characterised by potent cytotoxicity, e.g. swinholide A has an IC<sub>50</sub> of 0.04 and 0.03 µg/ml against KB and L1210 tumour cells *in vitro*.<sup>2a,d</sup> All of these marine macrolides have identical stereostructures,<sup>4</sup> determining their conformation and possibly the cytotoxic activity.<sup>2d</sup> As part of our synthetic studies towards swinholide A and misakinolide A, we now report the enantiocontrolled synthesis of the C<sub>19</sub>-C<sub>32</sub> / C<sub>17</sub>-C<sub>30</sub> segment 5.



Scheme 1

Scheme 2 summarises our strategy for the synthesis of the monomeric secoacid  $6^5$  for swinholide A (together with the secoacid 7 for misakinolide A), involving aldol-type disconnections at the C<sub>15</sub>-C<sub>16</sub> and C<sub>18</sub>-C<sub>19</sub> bonds to afford the key segments 5 and 8. Segment 5, containing the C<sub>19</sub>-C<sub>25</sub> stereopentad and the tetrahydropyran ring, should then be attainable using our general synthetic approach<sup>6a</sup> to such polypropionate systems. In this case, an *anti-anti* aldol reaction<sup>6b</sup> between the ethyl ketone (S)-10<sup>6</sup> and the chiral aldehyde 11 is required to control the C<sub>22</sub> and C<sub>23</sub> stereocentres in 9. The aldehyde component 11 should be available in turn from the cyclic acetal 12 by a suitable alkylation reaction at C<sub>27</sub>.



The synthesis of this C<sub>19</sub>-C<sub>32</sub> segment 5 starting from (E)-1,5-heptadien-4-ol  $(13)^7$  is shown in Scheme 3 and outlined below. Catalytic Sharpless asymmetric epoxidation<sup>8a</sup> of  $(\pm)$ -13 with kinetic resolution gave the (S, S, S) epoxide 14<sup>8b</sup> (96% ee by <sup>1</sup>H NMR analysis of the Mosher ester formed from (R)-(+)-MTPA) in 43% yield with 95% ds. Directed reductive opening of the epoxide 14 was achieved using Red-al<sup>\$\$\$9\$</sup> giving the 1,3-diol 15,<sup>10</sup>  $[\alpha]_D^{2D} = \pm 20.7^\circ$  (c 2.9, CHCl<sub>3</sub>). Ozonolysis of 15 in MeOH, followed by acidic workup and *O*-methylation, then gave the cyclic acetal 12 in 70% yield as a mixture of anomers, which were not separated. Treatment of 12 with allyltrimethylsilane in MeCN at -20 °C under Me<sub>3</sub>SiOTf catalysis<sup>11</sup> led, via kinetically controlled axial attack on the oxonium ion, to the rapid (< 2 min) and clean formation of the *trans*-substituted tetrahydropyran 16,<sup>10</sup>  $[\alpha]_D^{2D} = -62.3^\circ$  (c 3.8, CHCl<sub>3</sub>), in 96% yield with  $\ge 97\%$  ds. <sup>1</sup>H NMR decoupling and NOE difference experiments on 16 confirmed the relative stereochemistry at C<sub>27</sub> and suggested a preferred chair conformation with the allyl group axially disposed. A similar chair conformation is found for the tetrahydropyran containing segments of swinholide A.<sup>2b,d</sup> Ozonolysis of 16 then gave the corresponding aldehyde, which underwent a stereoselective Wittig reaction<sup>12</sup> to give the required (*E*)-enal 11,  $[\alpha]_D^{20} = -5.7^\circ$  (*c* 1.0, CHCl<sub>3</sub>), in preparation for the forthcoming aldol chain-extension.

Using our standard conditions with equimolar amounts of the two reactants,<sup>6b</sup> the key *anti*-selective boron aldol reaction between (S)-10 and the aldehyde 11 proceeded well. A high level of substrate-based stereocontrol at the C<sub>22</sub> and C<sub>23</sub> centres was achieved from the *E*-dicyclohexylenol borinate 17,<sup>6b</sup> giving the *anti-anti* (AA) isomer 9,<sup>10</sup> [ $\alpha$ ]<sub>D</sub><sup>20</sup> = -18.4° (c 2.0, CHCl<sub>3</sub>), in 84% yield with ≥97% ds (no other aldol isomers detected). This was followed by introduction of the C<sub>21</sub> stereocentre using reduction<sup>13</sup> with Me4BH(OAc)<sub>3</sub> to give the *anti*-1,3-diol 18, [ $\alpha$ ]<sub>D</sub><sup>20</sup> = +1.7° (c 1.8, CHCl<sub>3</sub>), with ≥97% ds, which was converted to its di-*tert*butylsilylene derivative 19 in 72% overall yield.

The remaining stereocentre at C<sub>24</sub> was installed by a hydroboration reaction on **19**, again relying on substrate-based<sup>14</sup> stereocontrol. Use of thexylborane gave, after oxidation, the alcohol **20**,  $[\alpha]_D^{20} = -45.3^\circ$ 

(c 2.2, CHCl<sub>3</sub>), in 74% yield with  $\geq$  97% ds. The surplus secondary hydroxyl group at C<sub>25</sub> was then efficiently removed by reduction<sup>15</sup> of the derived thiocarbonylimidazolide with <sup>n</sup>Bu<sub>3</sub>SnH, as in 20  $\rightarrow$  21 (80%). Finally, hydrogenolysis of the benzyl ether in 21 and subsequent Swern oxidation of 22 gave the desired aldehyde 5,  $[\alpha]_D^{20} = -75.9^\circ$  (c 1.3, CHCl<sub>3</sub>), in 90% overall yield. The assigned structure was verified using <sup>1</sup>H NMR (COSY, NOE).<sup>10</sup>

This completes a synthesis of a common  $C_{19}-C_{32} / C_{17}-C_{30}$  segment 5 for swinholide A and misakinolide A (15 steps from (±)-13, in 6% overall yield and 75% diastereoselectivity), using a combination of cyclic and acyclic stereocontrol strategies to set up seven of the eight stereogenic centres. In summary, this relies on a single *reagent*-controlled reaction, the Sharpless epoxidation  $13 \rightarrow 14$ , and a series of *substrate*-controlled reaction,  $12 \rightarrow 16$ , (*ii*) the boron-mediated aldol reaction,  $17 + 11 \rightarrow 9$ , (*iii*) the ketone reduction,  $9 \rightarrow 18$ , and (*iv*) the alkene hydroboration,  $19 \rightarrow 20$ . Studies towards the elaboration of aldehyde 5 into the antitumour macrolides swinholide A and misakinolide A are underway.



**Scheme 3** (a) (+)-DIPT (15 mol %), Ti(O<sup>i</sup>Pr)<sub>4</sub> (10 mol %), <sup>1</sup>BuOOH (50 mol %), 4Å sieves, CH<sub>2</sub>Cl<sub>2</sub>, -25 °C, 20 h; Me<sub>2</sub>S, 20 °C, 16 h; (b) Red-al<sup>®</sup>, THF, 20 °C, 18 h; (c) O<sub>3</sub>, MeOH, -20 °C, 10 min; Me<sub>2</sub>S, 20 °C; 1 *M* HCl(aq), 3 h; (d) NaH, MeI, THF, 20 °C, 6 h; (e) H<sub>2</sub>C=CHCH<sub>2</sub>SiMe<sub>3</sub>, Me<sub>3</sub>SiOTf (10 mol %), MeCN, -20 °C, 2 min; (f) O<sub>3</sub>, 3:1 CH<sub>2</sub>Cl<sub>2</sub>/MeOH, NaHCO<sub>3</sub>(s), -78 °C, 10 min; Me<sub>2</sub>S, 20 °C; (g) Ph<sub>3</sub>P=C(Me)CHO, PhH, reflux, 18 h; (h) (c-C<sub>6</sub>H<sub>11</sub>)<sub>2</sub>BCl, Et<sub>3</sub>N, Et<sub>2</sub>O, 0 °C, 2 h; 11, -78  $\rightarrow$  -20 °C, 14 h; H<sub>2</sub>O<sub>2</sub>, pH7 buffer, MeOH, 0 °C, 1 h; (i) Me<sub>4</sub>NBH(OAc)<sub>3</sub>, 1:1 AcOH/MeCN, -20 °C, 19 h; (j) <sup>1</sup>Bu<sub>2</sub>Si(OTf)<sub>2</sub>, 2,6-lutidine, CH<sub>2</sub>Cl<sub>2</sub>, 20 °C, 17 h; (k) thexylborane, THF, 20 °C, 3 h; H<sub>2</sub>O<sub>2</sub>/NaOH, 20°C, 1 h; (i) (imid)<sub>2</sub>C=S, THF, 60 °C, 16 h; (m) <sup>n</sup>Bu<sub>3</sub>SnH, PhMe, reflux, 50 min; (n) H<sub>2</sub>, 10% Pd/C, EtOH, 20 °C, 5 h; (o) (COCl)<sub>2</sub>, DMSO, CH<sub>2</sub>Cl<sub>2</sub>, -78 °C, 1 h; Et<sub>3</sub>N, -78  $\rightarrow$  -25 °C, 30 min.

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- All new compounds gave spectroscopic data in agreement with the assigned structures. 16 had <sup>1</sup>H NMR δ (CDCl<sub>3</sub>, 400 MHz) 5.75 (1H, dddd, J = 17.1, 10.0, 7.1, 7.0 Hz, CH=CH<sub>2</sub>), 5.05 (1H, d, J = 17.1 Hz, CH=CH<sub>A</sub>H<sub>B</sub>), 5.03 (1H, d, J = 10.0 Hz, CH=CH<sub>A</sub>H<sub>B</sub>), 4.05 (1H, m, H<sub>27</sub>), 3.73 (1H, dqd, J = 9.4, 6.2, 2.9 Hz, H<sub>31</sub>), 3.50 (1H, m, H<sub>29</sub>), 3.30 (3H, s, OMe), 2.43 (1H, m, CHAHBCH=C), 2.19 (1H, m, CHAHBCH=C), 1.95 (1H, m, H<sub>30eq</sub>), 1.84 (1H, m, H<sub>28eq</sub>), 1.52 (1H, ddd, J = 12.9, 10.2, 5.4 Hz, H<sub>28ax</sub>), 1.19 (1H, buried m, H<sub>30ax</sub>), 1.18 (3H, d, J = 6.2 Hz, Me<sub>32</sub>); <sup>13</sup>C NMR  $\delta$  (CDCl<sub>3</sub>, 100 MHz) 135.0, 117.0, 72.9, 71.4, 65.1, 55.2, 38.4, 36.7, 33.7, 21.6; HRMS (CI, NH<sub>3</sub>) [M+H]<sup>+</sup> found 171.1385, C<sub>10</sub>H<sub>19</sub>O<sub>2</sub> requires 171.1385; 9 had <sup>1</sup>H NMR & (CDCl<sub>3</sub>, 400 MHz) 7.36-7.27 (5H, m, Ph), 5.45 (1H, dd, J = 6.8, 6.6 Hz, H<sub>25</sub>), 4.52 & 4.47  $(2H, AB_q, J_{AB}= 12.1 \text{ Hz}, CH_2Ph), 4.18 (1H, dd, J = 9.3, 2.7 \text{ Hz}, H_{23}), 4.05 (1H, m, H_{27}), 3.72 (1H, m, H_{31}), 3.68 (1H, H_{27}), 3.72 (1H, H_{27$ dd, J = 8.9, 8.7 Hz, H<sub>19A</sub>), 3.53 (1H, m, H<sub>29</sub>), 3.45 (1H, dd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.34 (3H, s, OMe), 3.10 (1H, dqd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.53 (1H, s, OMe), 3.10 (1H, dqd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.54 (3H, s, OMe), 3.10 (1H, dqd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.54 (3H, s, OMe), 3.10 (1H, dqd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.54 (3H, s, OMe), 3.10 (1H, dqd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.54 (3H, s, OMe), 3.10 (1H, dqd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.54 (3H, s, OMe), 3.10 (1H, dqd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.54 (3H, s, OMe), 3.10 (1H, dqd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.54 (3H, s, OMe), 3.10 (1H, dqd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.54 (3H, s, OMe), 3.10 (1H, dqd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.54 (3H, s, OMe), 3.10 (1H, dqd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.54 (3H, s, OMe), 3.10 (1H, dqd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.54 (3H, s, OMe), 3.10 (1H, dqd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.54 (3H, s, OMe), 3.10 (1H, dqd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.54 (3H, s, OMe), 3.10 (1H, dqd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.54 (3H, s, OMe), 3.10 (1H, dqd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.54 (3H, s, OMe), 3.10 (1H, dqd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.54 (3H, s, OMe), 3.10 (1H, dqd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.54 (3H, s, OMe), 3.10 (1H, dqd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.54 (3H, s, OMe), 3.10 (1H, dqd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.54 (3H, s, OMe), 3.10 (1H, dqd, J = 8.9, 5.0 Hz, H<sub>19B</sub>), 3.54 (3H, s, OMe), 3.10 (1H, dqd), 3.10 8.7, 7.0, 5.0 Hz, H<sub>20</sub>), 2.88 (1H, dq, J = 9.3, 7.1 Hz, H<sub>22</sub>), 2.62 (1H, s, OH), 2.45 (1H, m, H<sub>26A</sub>), 2.21 (1H, m, H<sub>26B</sub>), 1.99 (1H, m, H<sub>30eq</sub>), 1.85 (1H, m, H<sub>28eq</sub>), 1.63 (3H, s, C<sub>24</sub>Me), 1.57 (1H, ddd, J = 12.9, 10.3, 5.5 Hz, H<sub>28ax</sub>), 1.20 (1H, m,  $H_{30ax}$ ), 1.19 (3 $\hat{H}$ , d, J = 6.2 Hz, Me<sub>32</sub>), 1.06 (3H, d, J = 7.0 Hz, C<sub>20</sub>Me), 0.92 (3H, d, J = 7.1 Hz, C<sub>22</sub>Me); <sup>13</sup>C NMR 8 (CDCl3, 100 MHz) 217.2, 137.8, 136.1, 128.4, 127.7, 127.6, 125.6, 80.0, 73.3, 73.1, 72.3, 71.8, 65.2, 55.3, 49.5, 45.8, 38.6, 34.0, 30.5, 21.7, 13.7, 13.6, 11.0; HRMS (CI, NH<sub>3</sub>) [M+NH<sub>4</sub>]<sup>+</sup> found 436.3063, C<sub>25</sub>H<sub>42</sub>O<sub>5</sub>N requires 436.3063; 5 had <sup>1</sup>H NMR  $\delta$  (C<sub>6</sub>D<sub>6</sub>, 400 MHz) 9.97 (1H, d, J = 2.6 Hz, CHO), 4.31 (1H, dd, J = 9.1, 3.6 Hz, H<sub>21</sub>), 4.02 (1H, m, H<sub>27</sub>), 3.70 (1H, dd, J = 4.8, 4.6 Hz, H<sub>23</sub>), 3.60 (1H, m, H<sub>31</sub>), 3.38 (1H, m, H<sub>29</sub>), 3.19 (3H, s, OMe), 2.46 (1H, dqd, J = 9.1, 7.0, 100) 2.6 Hz, H<sub>20</sub>), 1.89 (2H, m, H<sub>22</sub>, H<sub>28eq</sub>), 1.84 (2H, m, H<sub>26A</sub>, H<sub>30eq</sub>), 1.74 (1H, m, H<sub>28ax</sub>), 1.61 (2H, m, H<sub>24</sub>, H<sub>25A</sub>), 1.51 (1H, m, H<sub>25B</sub>), 1.30 (1H, m, H<sub>30ax</sub>), 1.25 (3H, d, J = 6.4 Hz, Me<sub>32</sub>), 1.16 (19H, s + buried m, H<sub>26B</sub>, Bu), 0.95 (3H, d, J = 6.7 Hz,  $C_{24}$ Me), 0.91 (3H, d, J = 7.3 Hz,  $C_{22}$ Me), 0.79 (3H, d, J = 7.0 Hz,  $C_{20}$ Me); <sup>13</sup>C NMR  $\delta$  (C<sub>6</sub>D<sub>6</sub>, 100 MHz) 203.2, 82.3, 75.4, 73.6, 71.0, 64.8, 55.0, 49.3, 38.9, 38.5, 36.7, 35.6, 29.3, 28.3, 27.9, 26.9, 22.2, 22.0, 16.1, 13.9, 10.8; HRMS (CI, NH<sub>3</sub>) [M+H]<sup>+</sup> found 471.3501, C<sub>26</sub>H<sub>51</sub>O<sub>5</sub>Si requires 471.3506.
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